

## Lecture 2

### tRNA: Structure & Function

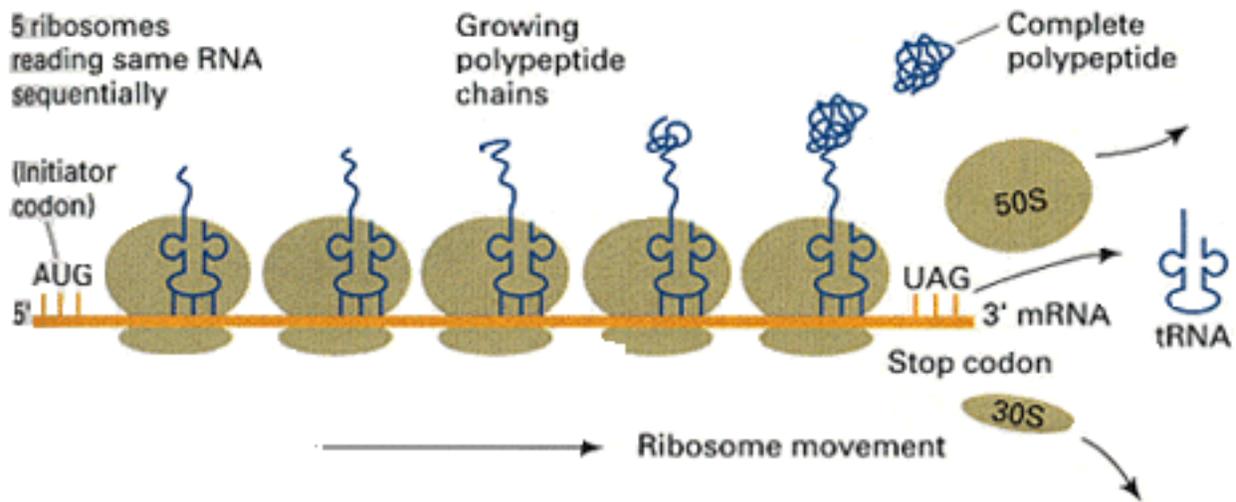
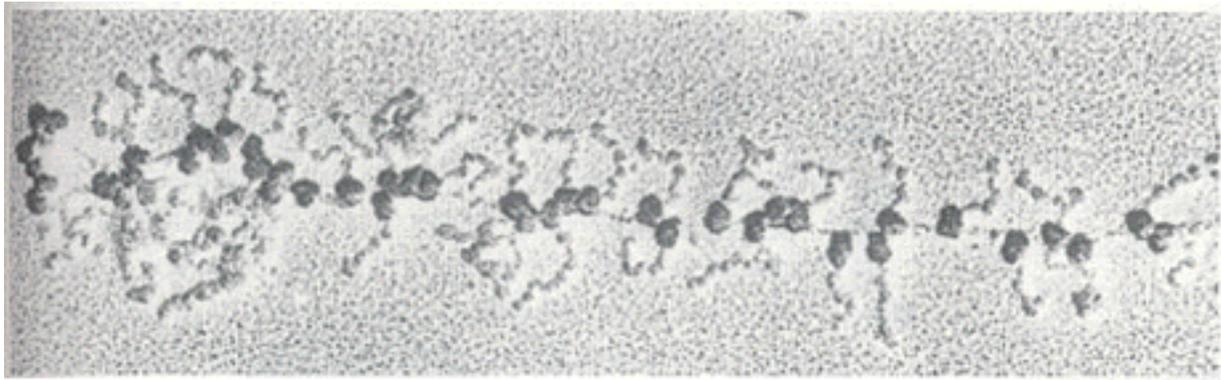
Major players in *protein synthesis*:  
mRNA, tRNA and the ribosome

<b>mRNA</b>	<b>Messenger RNA</b> , a copy of DNA blueprint of the <i>gene</i> to be expressed.	<b>Information</b>
<b>tRNA</b>	Aminoacyl <b>transfer RNA</b> , also called <i>anticodon</i> or <i>adaptor</i> molecule. One or more tRNAs for each amino acid.	<b>Supply</b>
<b>Ribosome</b>	A very large complex of several rRNAs (ribosomal RNA) and many protein molecules. Total molecular weight over 2 million dalton.	<b>Factory</b>

<b>Protein</b>	Polypeptide chain with sequence dictated by the mRNA sequence. Also called the <i>gene product</i> .	<b>Product</b>
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## Protein Synthesis

**Electronmicrograph of a so-called *polysome*: one mRNA strand (faint horizontal line) with many individual ribosomes attached (dark blobs). The newly synthesized *polypeptide* chains (*proteins*) can be seen as irregularly shaped extensions from the ribosomes:**



The bottom panel shows a schematic representation of the process in the upper panel.

Now let's magnify a ribosome to the size of a "Big Mac" (factor 10,000,000). At this magnification an *E. coli* bacterium would

be about 10 meters (or 30 feet) in diameter. You would be about 10,000 miles tall:

<b>Ribosome</b>	20 nm or 200 Å	<b>Big Mac</b> 20 cm	8 inches
<b>tRNA</b>	5 nm or 50 Å	5 cm	2 inches
<b>mRNA (900 bases)</b>	450 nm or 4,500 Å	450 cm	15 feet
<b>Extended (<i>unfolded</i>) protein (300 amino acids)</b>	90 nm or 900 Å	90 cm	3 feet
<b>Globular (<i>folded</i>) protein (300 amino acids)</b>	5 nm or 50 Å	5 cm	2 inches

1 nm (nanometer) is  $10^{-9}$  meters.  
1 Å (angstrom) is  $10^{-10}$  meters or 0.1 nm.

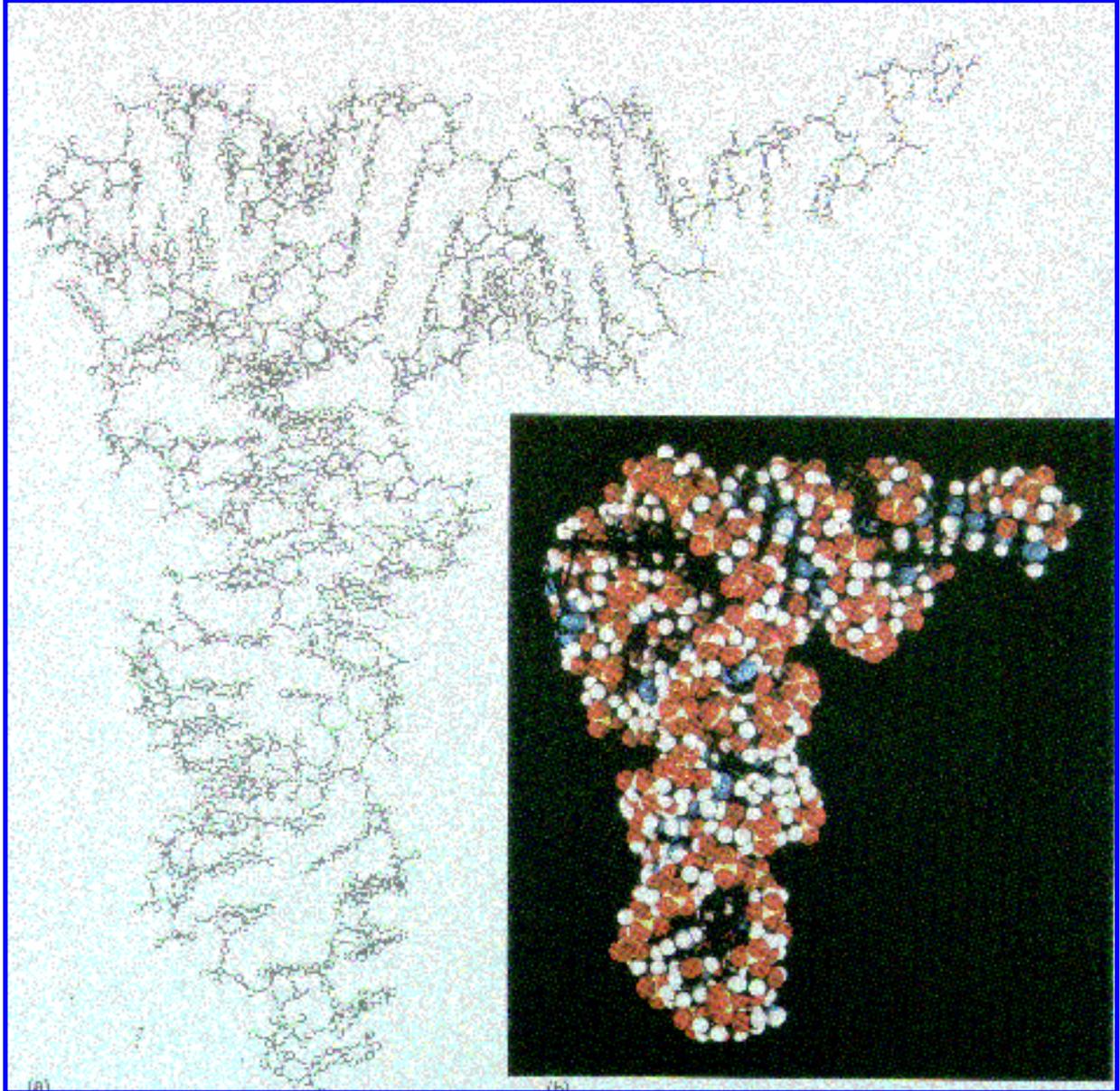
**[Next: tRNA Structure](#)**

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## Tertiary structure

- The tertiary structure of tRNA is best described as a compact "L" shape.
- The *anticodon* is a single-stranded loop at the bottom of the Figure which later base-pairs with the triplet codon
- The amino acid is attached to the terminal **A** on the upper right.
- The active sites (*anticodon* and *amino acid*) are maximally separated.
- As in proteins, the tertiary structure is dictated by the primary sequence.
- The tertiary structure is stabilized by base pairing and base stacking.
- Two areas (anticodon stem and acceptor stem) form double helix.



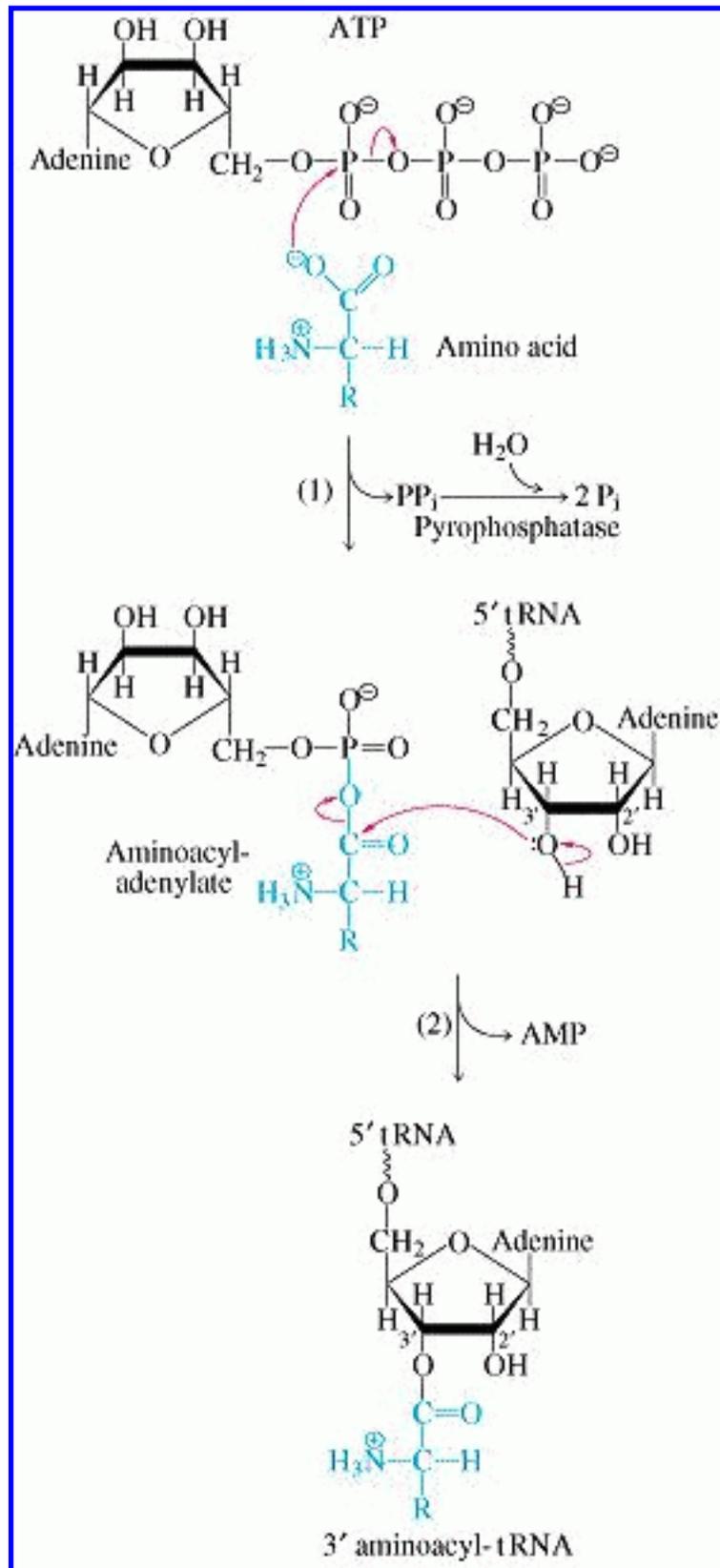
## [Next: tRNA Function](#)

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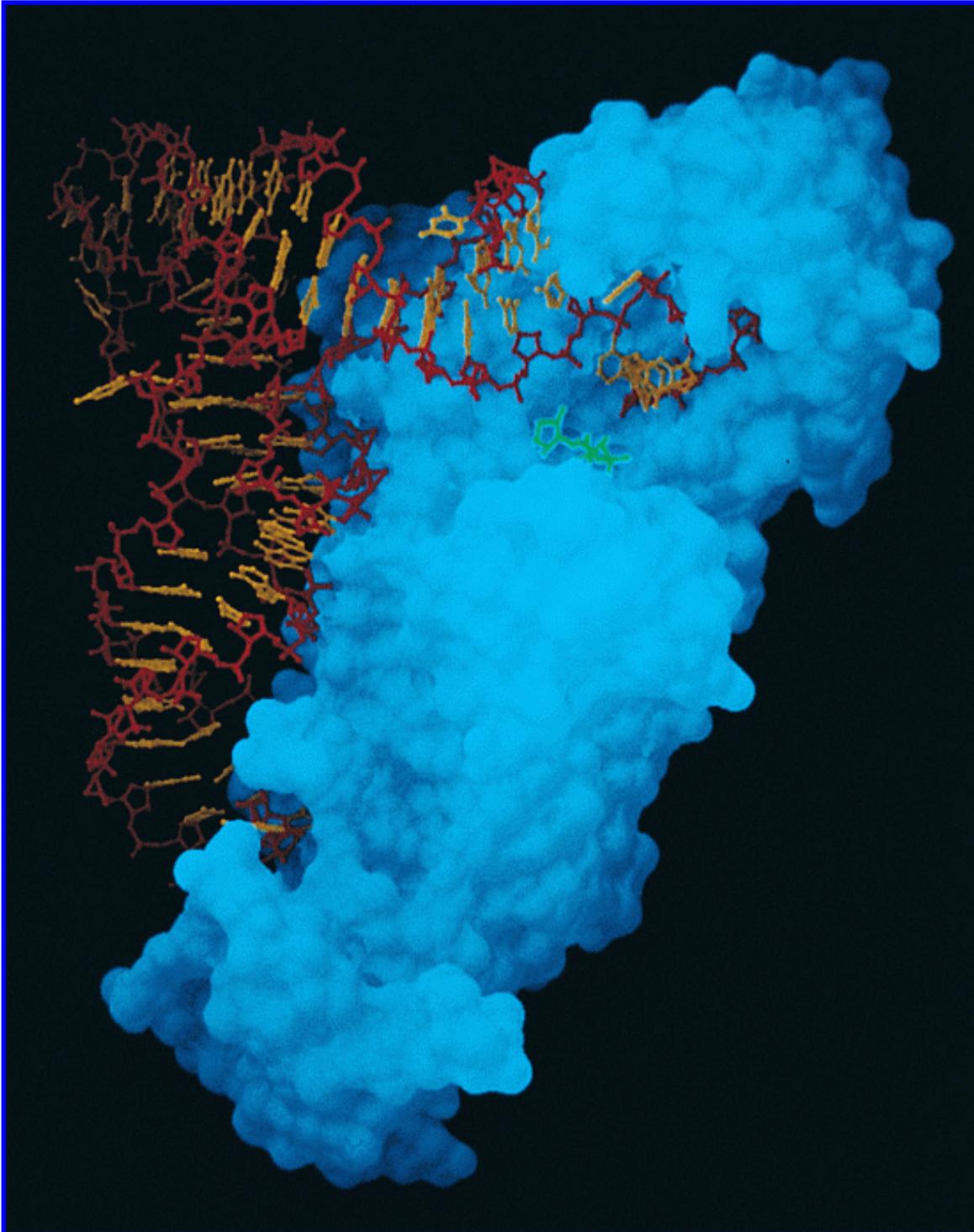
## tRNA Function: Synthetases

- Each tRNA is *charged* with the proper amino acid via a covalent ester bond at their 3' end by a family of enzymes called ***aminoacyl-tRNA synthetases***. Each enzyme must recognize ***both*** the tRNA specific for an amino acid and the corresponding amino acid. This energy-consuming process is ATP-dependent and results in the cleavage of **two** high-energy phosphate bonds (one in going from ATP to AMP + PP, and one for the cleavage of pyrophosphate into two inorganic phosphates):

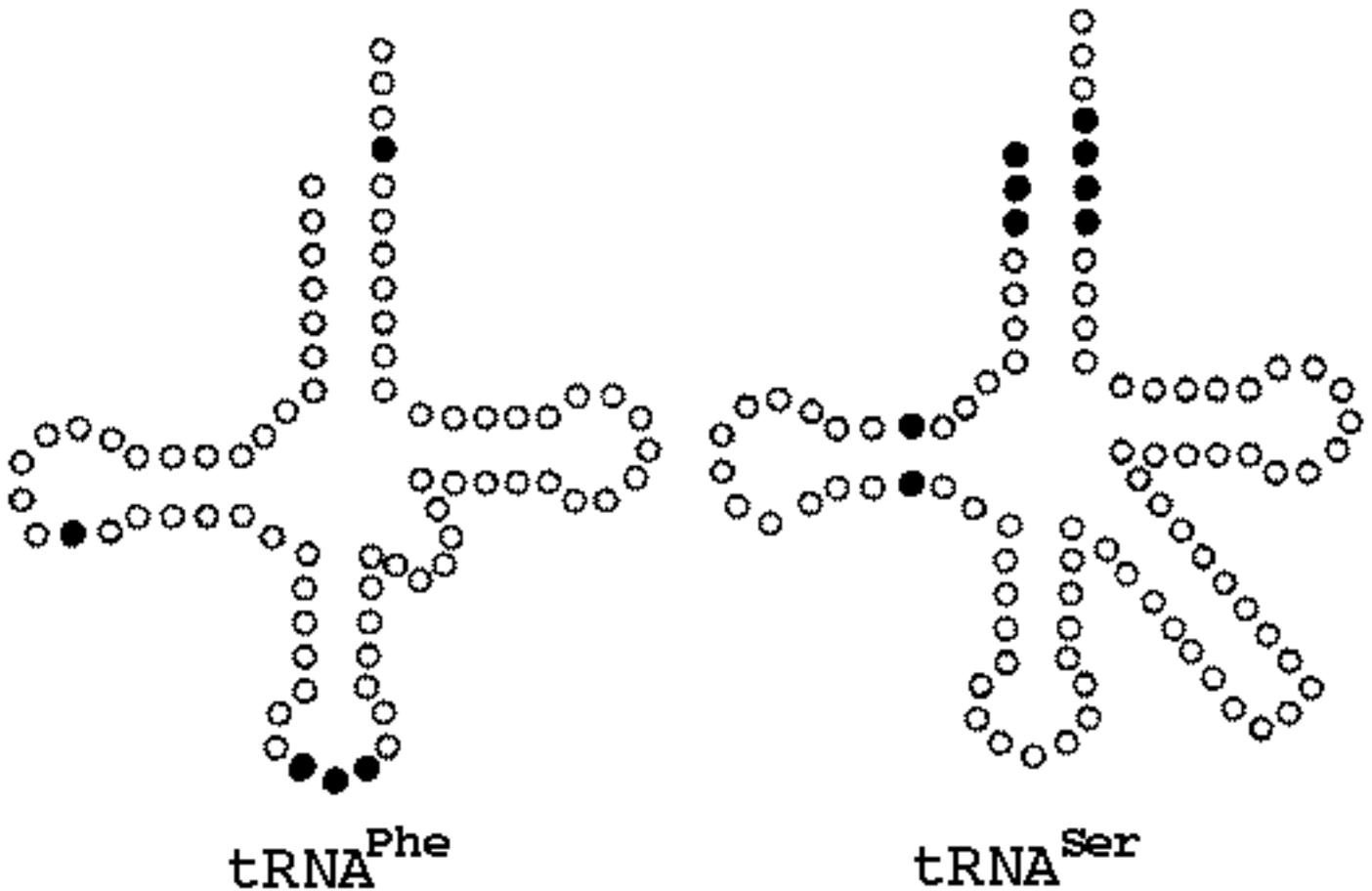


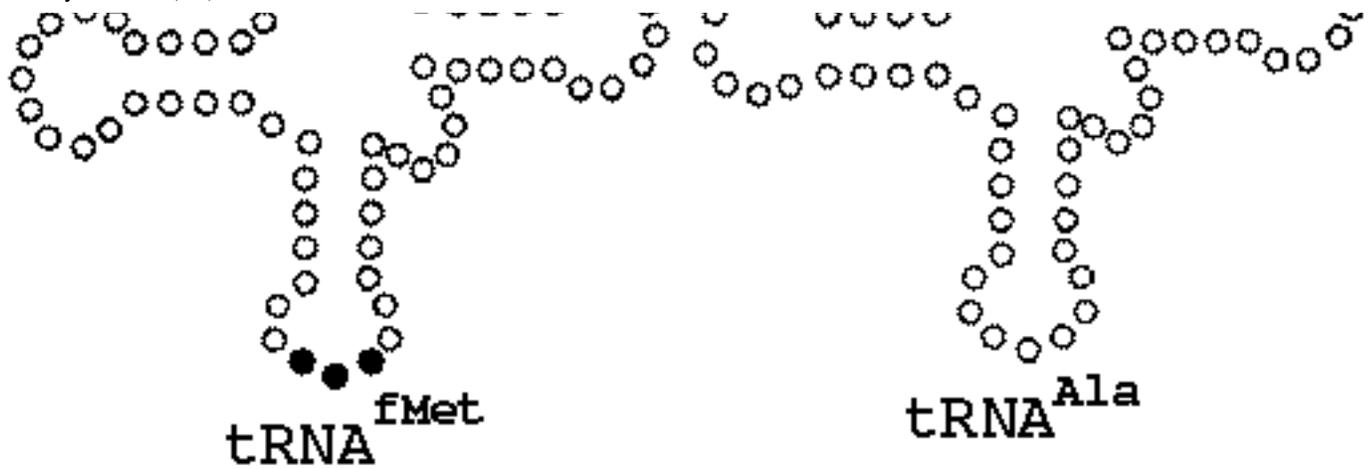
- There are 20 different aminoacyl-tRNA synthetases, one for each amino acid. Despite the fact that they all carry out very similar tasks, they vary greatly in size (40-100 kDalton).

- Since there are 61 amino acid codons, most tRNA synthetases must be able to recognize more than one type of tRNA (i.e. 6 codons for Arg). These tRNAs are called *cognate tRNAs* for that particular synthetase. This mapping is achieved through so-called *recognition domains* on the tRNA. tRNA shown with red backbone and yellow bases. tRNA synthetase shown as space-filling model in blue:



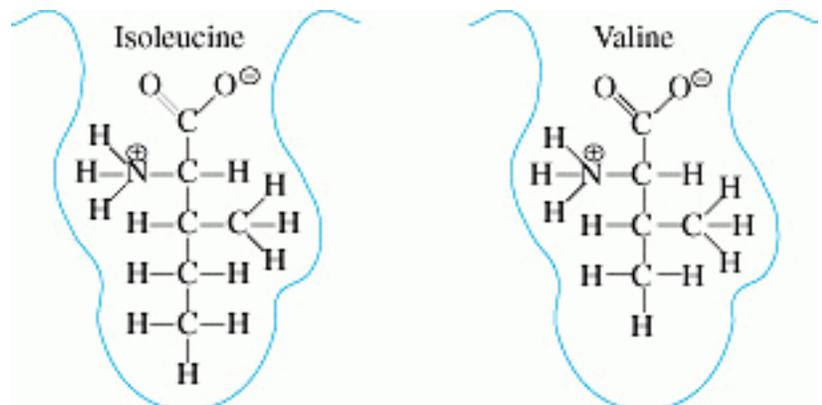
- The *recognition domain* includes unique sections of the acceptor stem *and/or* the anticodon (black dots):





## Accuracy & Proofreading

- The *accuracy* of charging tRNA with the proper amino acid is crucial because once charged, only the tRNA *anticodon* determines incorporation, *not* the attached amino acid.
- The error rate of charging is very low: 1 in 10,000. This is achieved by two means:
  - the amino acid specificity pocket in a specific synthetase will only bind amino acids similar in size and charge.
  - the synthetase also has



proofreading capability which, once a wrong aminoacyl-adenylate complex is formed (1st step), will hydrolyze the complex *before* it can be covalently attached to the tRNA (2nd step).

## Next: The Wobble Theory

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# The Wobble Theory

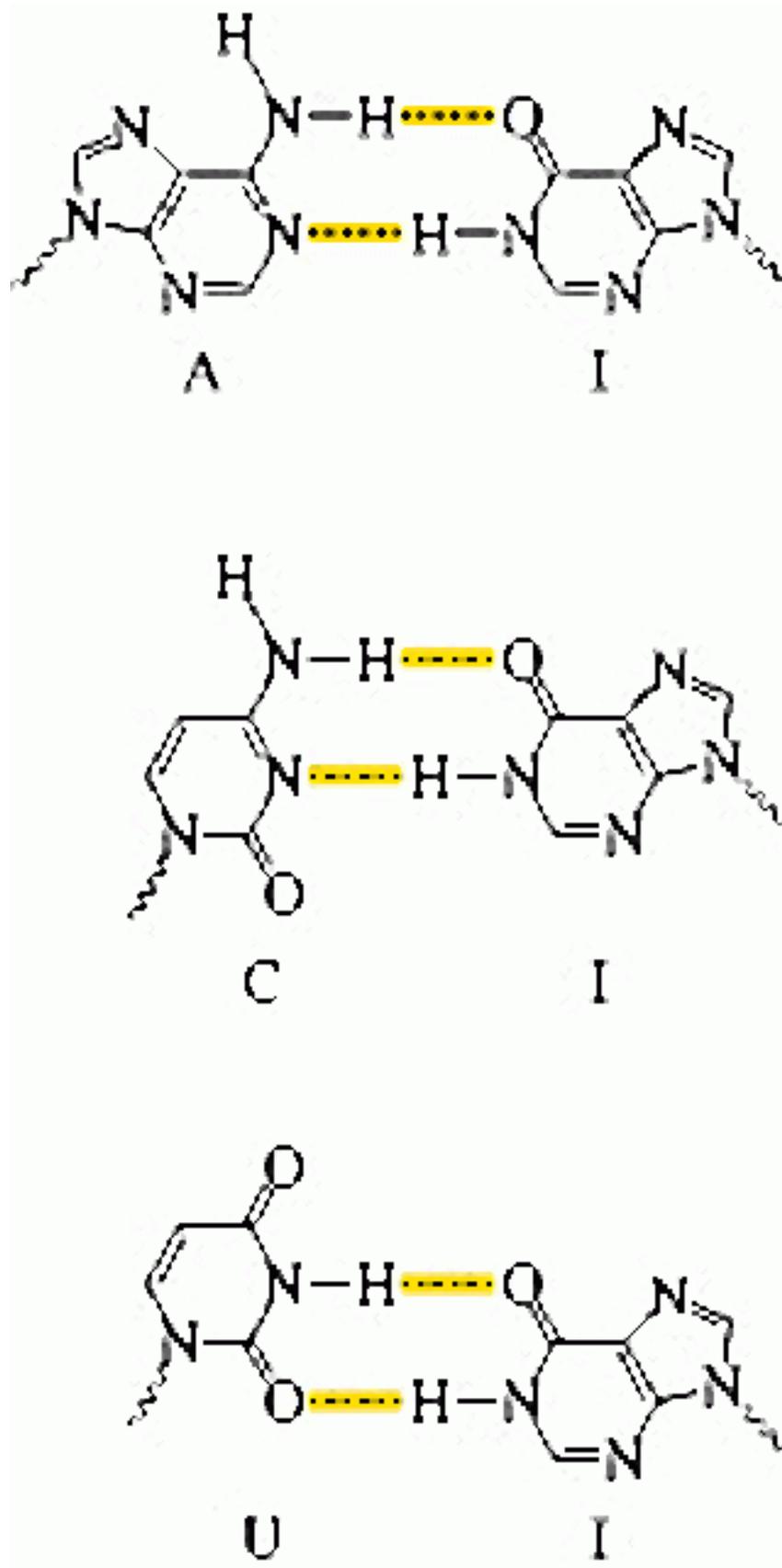
## Assumption:

Each tRNA, defined by its 3-base anticodon, pairs only with its complementary codon on the mRNA.

## Discrepancy:

tRNA<sup>Ala</sup> was found to bind to codons GCA, GCC and GCU.

Codon (5'→3')	GCA	GCC	GCU
Anticodon (3'→5')	CGU	CGG	CGA



## The answer:

The tRNA<sup>Ala</sup> anticodon is actually **CGI** which pairs with all three codons.

## Experimentally determined codon-anticodon pairing rules:

- The first two positions of the mRNA codon observe Watson-Crick base pairing rules (**A-U**, **C-G**)
- The third position exhibits *wobble*.
- Wobble occurs because the conformation of the tRNA anticodon loop permits flexibility at the first base of the anticodon.

5' anticodon base (tRNA)	3' codon base (mRNA)
<b>A</b> (not observed)	<b>U</b> (Watson-Crick)
<b>C</b>	<b>G</b> (Watson-Crick)
<b>G</b>	<b>C</b> or <b>U</b>
<b>U</b>	<b>A</b> or <b>G</b>

I	A or C or U
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**Caveat:** Since a single tRNA can respond to more than one codon, one tRNA could respond to codons for *two different* amino acids! This *would* lead to an ambiguous code. But since the Genetic Code is *unambiguous*, certain anticodons are disallowed.

## [Next: The Dangers of Wobble](#)

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# The Dangers of Wobble

Since a single tRNA can respond to more than one codon, one tRNA could *in theory* respond to codons for *two different* amino acids:

Example: [AGC](#) and [AGU](#) both code for [Ser](#). Their anticodon is **UCG** using [wobble rules](#). But anticodon **UCA** would also pair with **AGU** (**Ser**). This anticodon (**UCA**) would be converted [post-transcriptionally to UCI](#) (**Hey, that's us!**) which would recognize **AGC** and **AGU** as intended, but also [AGA](#) which codes for [Arg](#) -- not good, would result in an ambiguous Genetic Code!

One anticodon for serine is UCG:

Codon (5'→3')	AGC	AGU
Anticodon (3'→5')	UCG	UCG (wobble)
Amino acid	Ser	Ser

Now, in the following *hypothetical* example, anticodon UCA (*if it existed*) would also base-pair with codon AGU using wobble rules:

AGU	AGU	AGC	AGU	AGA
UCA =>	UCI (wobble)	UCI	UCI	UCI (wobble)
Ser	Ser	Ser	Ser	<b>Arg</b>

**But anticodon UCA would be converted post-transcriptionally to UCI which is able to bind three codons ([wobble rules](#)), including one for **Arg**! For this reason the UCA/UCI anticodon does not exist! And the codon AGA for Arg is actually covered by the anticodon UCU ([wobble](#))!**

**[Next: Supplemental Material](#)**

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# Supplemental Material

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## Molecular Graphics:

This exercise will let you get a better 3-dimensional picture of the interaction of a tRNA with its tRNA synthetase. You will be able to rotate, scale and otherwise manipulate the structure on your screen.

First, you'll need to obtain & install a **FREE** molecular graphics program called RASMOL on your PC:

[Click here](#) to obtain the MS Windows/NT version

or

[Click here](#) to obtain the Apple Macintosh version

Unpack the distribution, you should then be able to run the program RASWIN (PC) or RASMAC (MAC). [Click here for detailed help](#). If you get stuck or need help, please contact me ([Hudel](#)).

Second, you'll need to get a copy of the atomic coordinates from the Protein Data Bank:

[PDB entry 1GTR](#)

Save this rather large file (0.5 MBytes) with the "Save full entry to disk" option. Save the file as "**1GTR.pdb**". This file contains the complete atomic coordinates of GLUTAMINYL-tRNA SYNTHETASE COMPLEXED WITH tRNA AND ATP.

Now, run the program RASWIN/RASMAC and go to the "**Open...**" option under the "**File**" Menu. Select the file you just saved (**1GTR.pdb**). Then select the "**Backbone**" option under the "**Display**" menu. Your display should show something like [this](#). For a description of the program, check the "**User Manual**" under the "**Help**" Menu.

**Have fun!**

**[Next: Summary](#)**

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# Summary

## Features of tRNAs

- tRNAs are the *adaptor molecules* that translate from nucleic acid triplet to amino acid.
- tRNAs are *charged* with amino acids by enzymes called aminoacyl-tRNA synthetases in an energy-dependent 2-step process.
- There are 20 specific synthetases, one for each amino acid.
- Since most amino acids have more than one possible codon, there are more than 20 different tRNAs (40 in *E. coli*).
- Not all 61 codons have a specific tRNA due to *wobble* in the 3rd codon position.
- tRNA synthetases highly specific and have *proofreading* capabilities, achieving a better than 1:10,000 error rate.
- The accuracy of charging tRNA with the proper amino acid is *crucial* because once charged, only the tRNA *anticodon* determines incorporation into the growing polypeptide chain, *not* the attached amino acid.

### Next Lecture: The Ribosome, rRNA and mRNA

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